

A Bayesian Network Approach To Building Gene Regulatory Networks

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Abstract

Continuous abiotic stresses (such as extreme temperatures, high winds and edaphic conditions) can have adverse effects on plant life. This can become a major constraint in crop production. In order to alleviate these problems, it is important to understand the cold stress mechanisms at the molecular and cellular levels, regarding the signaling pathways from cold perception to activation of gene expression. We took a system biology approach, aiming to build the gene regulatory network using the cold stress microarray data that have been obtained, to gain a better understanding of plant responses to cold stress at the molecular level. The results generated from this approach will be utilized to generate hypothesis which can be tested by other experimental approaches.

To discover the gene regulatory network in response to cold stress on plants, we have employed a new proposed reverse engineering method implemented in the "GeneNet" R package (Opgen-Rhein and Strimmer 2007). This method is a heuristic algorithm applying an extended graphical Gaussian model and dynamical correlation shrinkage estimators to the inference of partial directed causal networks from high-dimensional time series expression data. Here, we analyze the expression data of 3, 376 interesting genes collected at 7 different time points (0, 0.5, 1, 3, 6, 12, 24 hours) from a microarry experiment. The resulting network shows 150 significant edges connecting 45 nodes.

Background

Plants respond to various environmental stresses such as cold stress by activating the expression of a large number of genes through a series of signal transduction cascades. These signal transduction cascades, often times interact with each other, compose the signaling networks, which eventually govern the transcriptional regulation of expression of genes which are involved in protecting plants from cold damage. Thus understanding the molecular mechanisms of the signal transduction network will greatly help in plant's stress tolerance, which in turn, will have a big economic influence on agricultural business.

Methods and Materials

The GeneChip® microarray data was downloaded from publicly available database (http://www.arabidopsis.org/info/expression/ATGenExpress.jsp). Plant total RNA were extracted from leaves of 18-day old plants treated with cold (40 C) for the indicated time periods, according to the standard protocol. cRNA labeling, hybridization and scanning was described in Hannah et al (2006). The Arabidopsis ATH1 whole genome microarray from Affymetrix was used for this set of cold stress experiments. All the data analysis was performed in R. The raw data file (.cel) files from chip hybridization were read directly into R. Robust Multi-Chip Analysis (RMA) algorithm was used for chip background correction, normalization and to obtain gene expression estimates.

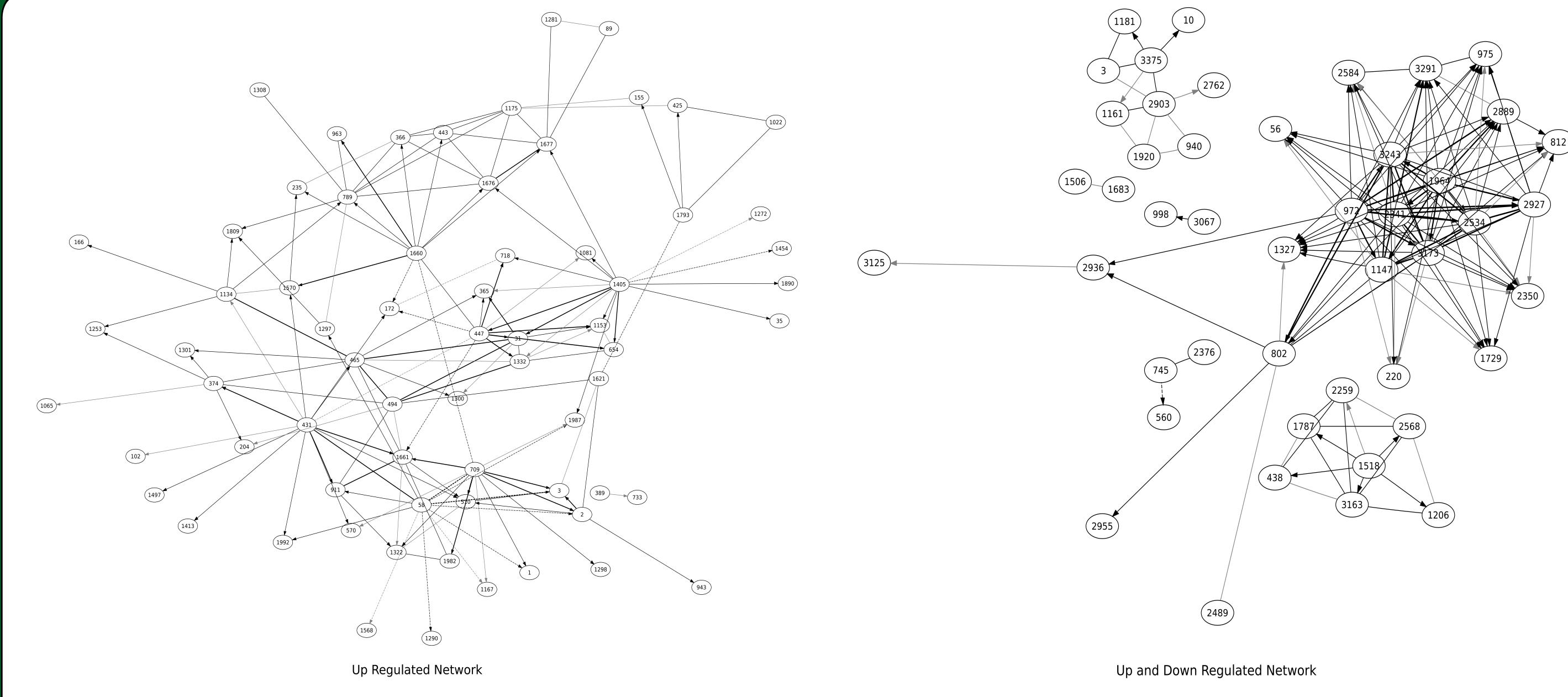
Local-Pooled Error (LPE) Test

Using the Local-Pooled Error (LPE) test we estimate the significance of each genes differential expression. LPE test is based on pooling errors within genes and between replicate arrays for genes in which expression values are similar (Jain et. al. 2003).

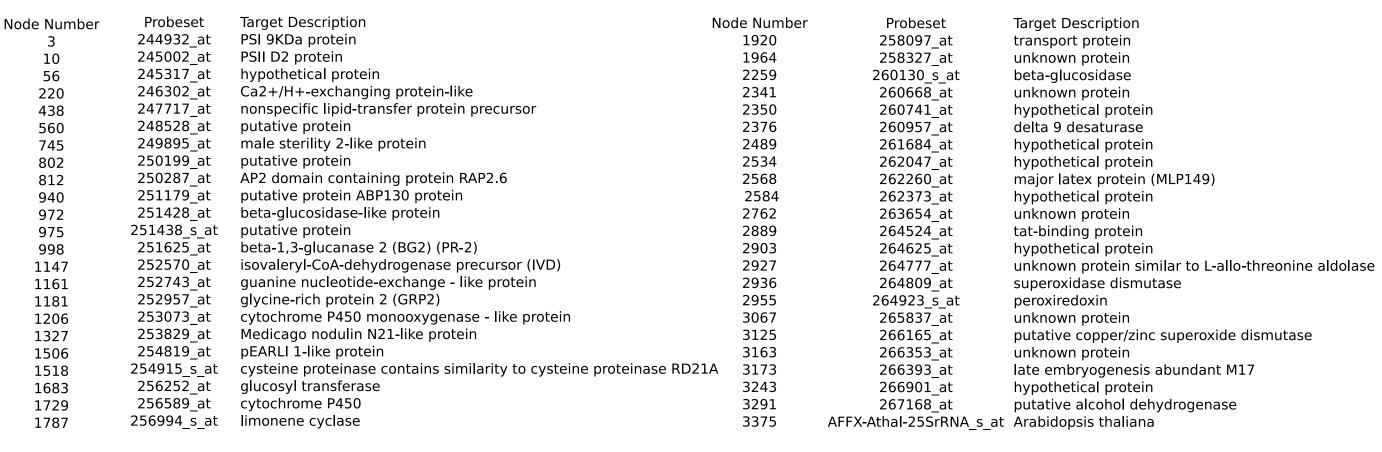
LPE is most useful when dealing with a low number of replicates (ie 2-3). This is one reason for using the LPE test rather than the traditional 2-sample t-statistics. Also, within-gene estimates of variance doe not provide a reliable hypothesis testing framework.

Procedures for LPE and how genes were selected to be used in GeneNet:

- 1. Evaluate baseline error distribution for each gene between replicates at each time point.
- 2. Calculate z-statistics for each gene. Here we allow positive z-scores denote up-regulation of genes and negative z-scores denote down-regulation.
- 3. Select genes with p-value less than or equal to 0.01
- 4. Once we have our up and down regulated genes with p-value less than or equal to 0.01, we further select genes with log2-fold change greater than or equal to 1.



Node Number	Probeset	Target Description	Node Number	Probeset	Target Description
1	244904_at	hypothetical protein	1065	257254_at	salicylic acid carboxyl methyltransferase
2	244977_at	cytochrome b/f	1081	257642_at	putative HLH DNA-binding protein
3	245015_at	large subunit of riblose-1,5-bisphosphate carboxylase/oxygenase	1134	258196_at	hypothetical protein
31	245397_at	auxin-responsive protein IAA1	1153	258399_at	early auxin-induced protein, IAA19
35	245422_at	putative protein	1167	258545_at	putative GTPase
58	245638_s_at	F5A9.10 unknown protein	1175	258675 at	putative nonspecific lipid-transfer protein
89	245928_s_at	vegetative storage protein Vsp1	1253	259391_s_at	delta 9 desaturase
102	246103_at	putative protein	1272	259583 at	hypothetical protein
155	246687_at	putative protein proline-rich protein APG	1281	259640 at	beta-glucosidase
166	246860_at	putative protein various predicted proteins	1290	259713 at	unknown protein similar to phosphate translocators
172	246888_at	putative protein	1297	259766_at	unknown protein
204	247252_at	unknown protein	1298	259767 s at	unknown protein
235	247463_at	embryo-specific protein - like embryo-specific protein 3	1300	259784 at	auxin-induced protein
365	248801_at	homeobox-leucine zipper protein-like	1301	259787 at	auxin-induced protein
366	248807_at	pectin methylesterase-like	1308	259871 at	nodulin-like protein
374	248888_at	potassium channel protein KAT1	1322	260007 at	unknown protein
389	249065_at	putative protein	1332	260152 at	putative IAA6 protein
425	249614_at	putative protein predicted proteins	1405	260957 at	delta 9 desaturase
431	249645_at	thionin Thi2.2	1413	261046 at	flavonol 3-o-glucosyltransferase
443	249813_at	acyltransferase	1454	261450 s at	O-methyltransferase
447	249895_at	male sterility 2-like protein	1497	262003 at	unknown protein
465	250012_x_at	auxin-induced protein-like	1568	262797 at	putative sugar transporter protein
494	250327 at	putative serine rich protein	1570	262819 at	putative cytochrome P450
510	250446_at	nucleoid DNA-binding protein cnd41	1621	263509 s at	hypothetical protein
570	251221 at	putative protein ER6 protein	1660	263979 at	En/Spm-like transposon protein
654	252204 at	putative protein	1661	263981_at	unknown protein
709	252957_at	glycine-rich protein 2 (GRP2)	1676	264146 at	hypothetical protein
718	253066 at	1-aminocyclopropane-1-carboxylate synthase - like protein	1677	264147_at	receptor-like protein glossy1 (gl1)
733	253203 at	arginine decarboxylase SPE2	1793	265441 at	unknown protein
789	253753 at	glycine-rich protein like glycine-rich protein	1809	265698 at	hypothetical protein predicted by genscan
911	254882 s at	putative protein various predicted reverse transcriptases/transposons		266606_at	putative AP2 domain transcription factor
943	255403 at	putative GH3-like protein	1982	267545 at	unknown protein
963	255782 at	transcription factor	1987	267595 at	putative glucanse
1022	256597 at	acidic ribosomal protein P2b (rpp2b)	1992	267644_s_at	unknown protein
		L	1332	20/044_3_at	unknown protein



GeneNet

GeneNet implements a statistical learning algorithm proceeding in two steps: (i) transform correlation network into a partial correlation network, which is an undirected graph displaying the linear associations, and (ii) convert the undirected graph into a partially directed graph by estimating pairwise log-ratios of standardized partial variances.

Consider a linear regression with Y as response and the set of vectors $\{X_1, \ldots, X_k, \ldots, X_K\}$ as covariates. The regression coefficient estimator of Y is defined by

$$eta_k^{\gamma} = \underbrace{ ilde{
ho}_{\gamma k}}_{\mathcal{A}} \underbrace{\sqrt{rac{SPV_{\gamma}}{SPV_k}}}_{\mathcal{B}} \underbrace{\sqrt{rac{\sigma_{\gamma}^2}{\sigma_k^2}}}_{\mathcal{C}}$$

where $\tilde{\rho}_{\gamma k}$ is the partial correlation between Y and X_k , and $SPV_{\gamma} = \tilde{\sigma}_{\gamma}^2/\sigma_{\gamma}^2$ called standardized partial variance.

- \mathcal{A} : Establish edges between nodes. An edge is drawn if $\mathcal{A} \neq 0$ determined by a multiple testing method (Efron 2004).
- \mathcal{B} : Determine the directionality of edges of a causal network. If $\log \mathcal{B} \neq 0$ evaluated by the multiple testing, edges are directed in a fashion that direction of arrow points from the node with larger standardized partial variance to the node with smaller standardized partial variance.

Summary and Future Research

We have built a gene regulatory network in plant cold stress response, based on the time point microarray data. We have identified a number of master regulatory genes which might play important roles in transducing cold stress signals to other genes. The network built in this work sets up a framework for bench scientists to design experiments to further test the importance of those genes, which might be important in cold stress response.

References

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Acknowledgements

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